



Models to study optimized combinations of physiological biotics for acne-prone skin

Gault Manon*; Bonnet Isabelle; Bertossi Eva; Andre-Frei Valérie
BASF Beauty Care Solutions France SAS.

Introduction:

Skin microbiota is involved in skin homeostasis and protects the skin against colonization by pathogenic microorganisms [1].

However, under certain conditions, perturbation of the skin microbial balance could occur and may lead to harmful conditions like **acne disorders**.

In acneic prone skin, **Cutibacterium acnes virulent** strains increase and cause **inflammation and redness**.

However, some skin “protectors” such as *Staphylococcus epidermidis* can protect the skin against the opportunistic pathogens [2]. This bacterium can notably use fermentation initiators (FIs) to produce short chain fatty acids (SCFAs) reducing the growth of *C. acnes* [3].

To better study and optimize this specific potential of *S. epidermidis* against *C. acnes*, a **bacterial and 3D microbiotic skin model** was developed to evaluate the **bacterial interactions and the effect of biotic ingredients**. To carry out these models, two bacteria were chosen: *S. epidermidis* strain ATCC 12228, a reference commensal strain [4] and *C. acnes* ATCC 6919, a bacterium isolated from acne lesions on human facial skin [5].

Materials & Methods:

Bacterial preparation

S. epidermidis strain ATCC 12228 was inoculated in Tryptic Soy Broth medium (TSB) at 37 °C for 24h under continuous stirring at 150 rpm in aerobic condition. *C. acnes* ATCC 6919 was inoculated in Schaedler medium at 37 °C for 72h without stirring in anaerobic condition.

Bacterial growth in monoculture model

Bacterial suspensions were prepared by adjusting the concentration of *S. epidermidis* at OD =0.05 at 600nm and at OD =0.3 for *C. acnes* in fresh medium diluted halfway with Phosphate Buffer Saline (PBS) in 96 well plates, with or without biotic ingredients (FI + SCFA) at 0.2 % and 2%. The bacterial growth was measured by densitometry (OD with a spectrophotometer) after 24h in aerobic condition for *S. epidermidis* or 72h in anaerobic condition for *C. acnes* (n=4).

Bacterial growth in coculture model

Mixt bacterial suspensions were prepared by adjusting the concentration of both bacteria at 1.10^6 in TSB medium. The coculture was incubated in 37°C for 72h without stirring in anaerobic condition, with or without biotic ingredients in 2% (FI + SCFA). To count *S. epidermidis*, serial dilutions were spread in TSB agar plates and incubated 24h in aerobic condition; To count *C. acnes*, serial dilutions were spread on TSB agar plates supplemented by furazolidone and incubated 72h in anaerobic condition (n=3).

Bacterial growth in coculture in 3D microbiotic skin model (RHE)

Keratinocytes (15 yo donor) were grown on the top of an insert for 4 days of submerged culture in DMEM/HAM-F12 medium with hydrocortisone 0.4µg/mL, insulin 4.16µg/mL, ascorbic acid 50µg/mL, EGF 10ng/mL and 5% of Bovine Calf Serum (BCS) before elevation at the air-liquid interface for 7 days in the same medium except for BCS at 1% and without EGF.

Reconstructed Human Epidermis (RHE) were infected by *S. epidermidis* or *C. acnes* separately or combined at 10^5 and 10^7 respectively. After 1 hour, samples were rinsed with PBS and then cultured up to 3 days post-infection. The efficiency of bacterial adhesion with biotic ingredient at 2% (FI + SCFA) was estimated using the colony forming unit assay at 1 hour post infection to evaluate adhesion and 72 hours post-infection for growth (n=3).

Acknowledgments:

We would like to thank the expert assistance of Mrs Marion Da Silva, Mr Sébastien Cadau and Mrs Aurélie Courtois.

References:

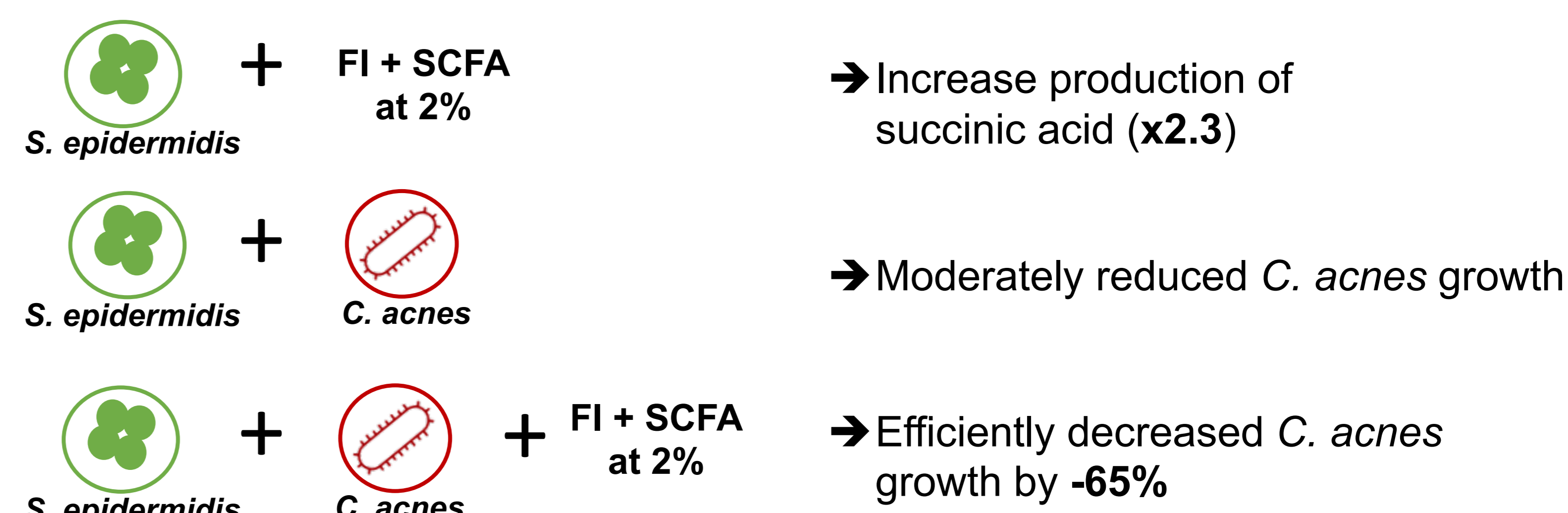
- Grice EA and Segre JA (2011) The skin microbiome. Nat Rev Microbiol, 9(4), 244-253.
- Wang Y, et al. (2014) *Staphylococcus epidermidis* in the human skin microbiome mediates fermentation to inhibit the growth of *Propionibacterium acnes*: implications of probiotics in acne vulgaris, Appl Microb Biotech 98:411–424
- Wang Y, et al. (2016) A Precision Microbiome Approach Using Sucrose for Selective Augmentation of *Staphylococcus epidermidis* Fermentation against *Propionibacterium acnes*. Int J Mol Sci. Nov 9;17(11):1870
- MacLea KS and Trachtenberga AM (2017) Complete Genome Sequence of *Staphylococcus epidermidis* ATCC 12228 Chromosome and Plasmids, Generated by Long-Read Sequencing. Genome Announc Sep; 5(36)
- McDowell A, et al. (2017) Proposal to reclassify *Propionibacterium acnes* type I as *Propionibacterium acnes* subsp. *Acnes* subsp. nov. and *Propionibacterium acnes* type II as *Propionibacterium acnes* subsp. *Defendens* subsp. nov. Int J Syst Evol Microbiol. Nov;67(11):4880

Results & Discussion:

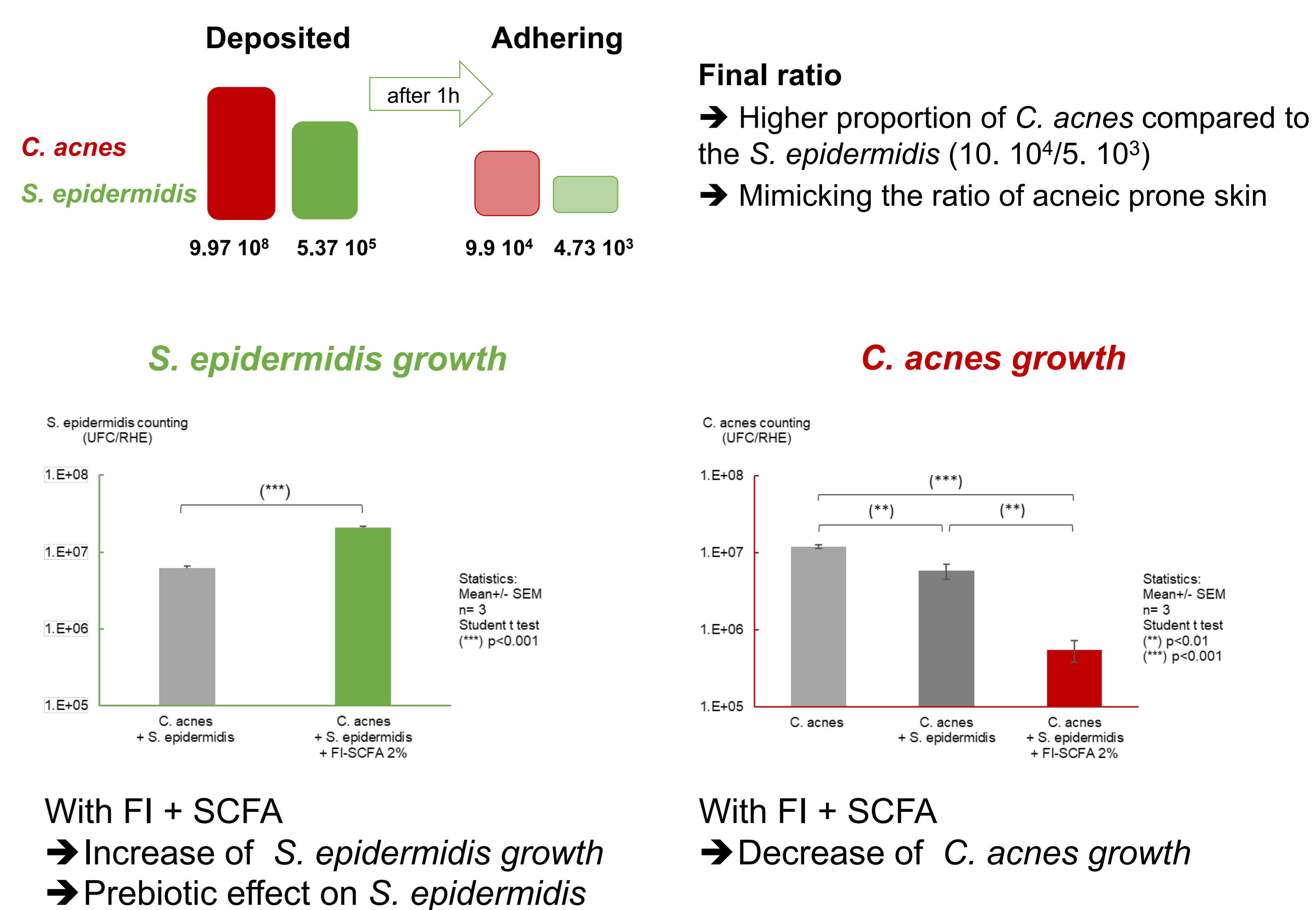
Monoculture



Coculture



3D microbiotic epidermal model



Conclusions:

From a simple model in monoculture to a more complex model of a co-culture of bacteria infecting a RHE, the **natural protective effect of *S. epidermidis* inhibiting *C. acnes* growth has been demonstrated**. All these models have also allowed to **select and develop biotic ingredients** improving those beneficial effects.

Studying the interactions between commensals and opportunistic pathogens helped us to design actives **inspired by the natural ecology of the skin microbiota** to promote the natural mechanisms of skin defenses.